1 Penicillin Detection by Tobacco Mosaic Virus-Assisted

2 Colorimetric Biosensors

- 3 Claudia Koch¹, Arshak Poghossian^{2,3}, Michael J. Schöning^{2,3} and Christina Wege^{1*}
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- ⁵ ¹Institute of Biomaterials and Biomolecular Systems, University of Stuttgart, 70569 Stuttgart,
- 6 Germany
- 7 ²Institute of Nano- and Biotechnologies, FH Aachen, Campus Jülich, 52428 Jülich, Germany
- 8 ³Peter Grünberg Institute (PGI-8), Forschungszentrum Jülich GmbH, 52525 Jülich, Germany
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10 Supplementary

- 11 Supplementary Figures:
- 12 Figure S1: Color changes of pH indicators between pH 1 and pH 12.
- 13 Figure S2: Absorption spectra of pH indicators.
- 14 Figure S3: Absorption changes in mixtures of penicillinase, penicillin (penG) and pH indicators.
- 15 Figure S4: Functionalization of TMV_{cys} particles and conjugation of streptavidin ([SA]) to penicillinase (Pen).
- 16 Figure S5: Detection and degradation of different β-lactam antibiotics by B. cereus penicillinase.
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22 Figure S1: Color changes of pH indicators between pH 1 and pH 12.

- 23 Test solutions with pH values ranging from pH 1 to pH 12 were supplemented and mixed with the distinct dyes
- 24 indicated at the left (for details, refer to the main text).
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27 Figure S2: Absorption spectra of pH indicators.

Absorption spectra (300 nm - 700 nm) of the halochromic dyes indicated above, incubated at pH values from
pH 1 to pH 12 (see Figure S1). The wavelength with maximum absorption change over the tested pH range was
determined for every individual dye: phenol red: 558 nm, bromcresol purple: 588 nm, methyl red: 516 nm,
bromthymol blue: 592 nm, cresol red: 572 nm, phenolphthalein: 555 nm.

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35 Figure S3: Absorption changes in mixtures of penicillinase, penicillin (penG) and pH indicators.

Variable enzyme amounts (5 ng \approx 12.5 μ U; 10 ng \approx 25 μ U; 15 ng \approx 37.5 μ U; 20 ng \approx 50 μ U; 25 ng \approx 62.5 μ U) blended with 5 mM penG or 10 mM penG, respectively, were mixed with pH indicator solutions. The absorption of each mixture was measured at the pH indicator-specific wavelength (see Figure S2) and the absorption change (OD/min) of each sample determined.

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45 Figure S4: Functionalization of TMV_{cys} particles and conjugation of streptavidin ([SA]) to penicillinase (Pen).

46 A) Denaturing SDS-PAGE reveals a selective coupling of maleimide-PEG₁₁-biotin (Bio) linker molecules to coat 47 protein units (CP_{Cys}) of TMV_{Cys} particles, resulting in a distinguishable mobility shift (CP_{Cys}/Bio). Image evaluation 48 by ImageJ confirmed a biotinylation efficiency of around 95 % (see main text for details). B) The enzyme 49 penicillinase from *B. cereus* is a mixture of two types of β -lactamases (E.C.3.5.2.6). To enable bioaffinity binding 50 of the enzymes to TMV_{Cys}/Bio, penicillinase molecules were conjugated with streptavidin [SA] using a LYNX 51 Rapid Streptavidin Antibody Conjugation Kit®, resulting in [SA]-Pen. To verify conjugation of [SA] and Pen, 52 samples were analyzed by 12 % SDS-PAGE and stained with Coomassie Brilliant Blue R250. Successful 53 conjugation of one or more streptavidin tetramers per penicillinase was confirmed by appearance of new band 54 signals corresponding to $[SA]_n$ -Pen, accompanied by weaker signals for bare β -lactamase.

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58 Figure S5: Detection and degradation of different β-lactam antibiotics by *B. cereus* penicillinase.

50 mU or 100 mU penicillinase were blended with analyte solutions containing variable concentrations (0 mM –

50 mM) of one out of six different antibiotics (penicillin G, tricarcillin, carbenicillin, ampicillin, cloxacillin,

61 cefotaxim). Enzymatic degradation of the antibiotic was detected by a color change of the pH indicator.

62 A) Photograph showing the color of mixtures containing penicillinase, antibiotics and bromcresol purple after

63 15 min incubation. B) The absorption of each mixture was measured at the pH indicator-specific wavelength

64 (λ =588 nm) and the absorption change (OD/min) of each sample determined.